Forum Review

The Pulmonary Biology of Isoprostanes

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ABSTRACT

Isoprostanes were first recognized as convenient markers of oxidative stress, but their powerful effects on a variety of cell functions are now also being increasingly appreciated. This is particularly true of the lung, which is comprised of a wide variety of different cell types (smooth muscle, innervation, epithelium, lymphatics, etc.), all of which have been shown to respond to exogenously applied isoprostanes. In this review, we summarize these biological responses in the lung, and also consider the roles that isoprostanes might play in a range of pulmonary clinical disorders. *Antioxid. Redox Signal.* 7, 244–255.

INTRODUCTION

As OUTLINED ELSEWHERE in this compendium, isoprostanes are found in abundance in a diverse variety of experimental and clinical conditions that have at least one feature in common: production/accumulation of free radicals and reactive oxygen species. For this reason, they have been frequently used as markers of oxidative stress. However, it is becoming increasingly clear that these molecules are not merely breakdown products nor convenient and inert markers, but in fact also exert a wide variety of biological responses. This is particularly true of the lung, in which it seems every type of cell present exhibits one or more physiologically relevant changes in function in response to isoprostanes (Fig. 1). The intent of this review is to summarize the current understanding of the effects of isoprostanes on pulmonary biology and pathophysiology.

Before doing so, however, it would be helpful to discuss briefly the current ideas regarding the receptors through which isoprostanes are believed to act. Even some of the very earliest studies pertaining to isoprostanes examined this question, finding the responses to be sensitive to several structurally different agents with relatively potent and selective inhibitory actions against thromboxane receptors (TP receptors) (7, 79, 109). Since then, numerous pharmacological studies of isoprostane-mediated responses have demonstrated

an involvement of prostanoid receptors, usually those of the TP subtype, although more recent studies have also uncovered evidence for the involvement of prostaglandin (PG) E₂selective (17, 24, 52, 100, 116) and PGF₂₀-selective (64, 116) prostanoid receptors (EP and FP, respectively). Isoprostanes and prostanoids share a similar structure, differing only in the orientation of their aliphatic side chains; thus, it is reasonable to assume that they are capable of forming similar binding interactions with receptor active-site residues. Indeed, one recent study (115) has shed some light on the structural features of PGE, that are required for binding to the EP, prostanoid receptor, these include the hydroxyl groups at the 11 and 15 positions, the terminal carboxyl group, and the omega-tail (115), all of which are found in most of the 15-isoprostanes. The ketone group at the 9 position, which is unique to the Ering prostanoids and isoprostanes, is also involved (115), which might account for the greater potency of the E-ring isoprostanes in many of the biological responses that we have studied (summarized below). Another study that examined ligand interactions at the TP receptor found that the substituents at positions 11 and 15 are particularly important determinants of reactivity, and that removal of the double bonds at positions 5,6 or 13,14 abolishes ligand activity (67). On the other hand, others argue for the existence of a unique class of isoprostane-selective receptors (26-28) (summarized elsewhere in this compendium).

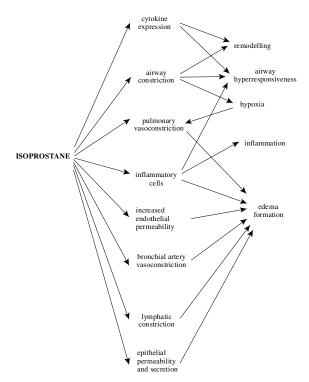


FIG. 1. Isoprostanes orchestrate a myriad of effects on most (all?) of the cell types found within the lungs. Altogether, these effects may account for many of the manifestations of asthma.

AIRWAY SMOOTH MUSCLE (ASM)

Direct effects on mechanical activity

Pharmacological studies of human ASM show that TP receptors predominate (5, 82); as such, it is not surprising that isoprostanes evoke substantial contractions that are sensitive to the TP-receptor antagonist ICI 192605 (55, 61). 8-iso PGE_2 is the most potent and powerful, acting at submicromolar concentrations to evoke much larger contractions than cholinergic agonists (the primary excitatory neurotransmitter in ASM). 8-iso $PGF_{1\alpha}$ and 8-iso $PGF_{2\alpha}$ are also somewhat excitatory in this preparation, but require ~10-fold higher concentrations to exert the same response as 8-iso PGE_2 . Other isoprostanes that we have tested elicit little or no increase in tone in human ASM, including 8-iso $PGF_{1\beta}$, 8-iso $PGF_{2\beta}$, 8-iso $PGF_{3\alpha}$ (55), and several dinor and tetranor derivatives of 8-iso $PGF_{2\alpha}$ (unpublished observations).

Bovine ASM also demonstrates contractile responses to 8iso PGE₂ (whereas the F-ring isoprostanes are ineffective). These appear to involve excitatory non-TP prostanoid receptors (EP or FP?), because the thromboxane agonist U46619 evokes relatively small contractions in these tissues only at concentrations in the micromolar range (orders of magnitude higher than needed for stimulation of TP receptors).

In the dog, TP receptors are largely absent in the larger airways and increase in number as one progresses down the airway tree (51); likewise, porcine airways appear to lack TP receptors, because the TP agonist U46619 does not evoke

constriction in this tissue (however, see contradictory findings summarized under *Electrophysiology*). Consistent with this, none of the isoprostanes that we have tested evoke a constrictor response in dog trachea (55) nor in pig airways (17), whereas moderate contractions are seen in dog bronchi (55).

Inhibitory EP receptors, however, are prevalent in ASM of all species: these are of the EP₂ subtype in human (83), canine (17), and murine (104) airways. Not surprisingly, then, isoprostanes can evoke a relaxant response in canine and porcine airways (17), as well as human airway tissues pretreated with TP-receptor blockers to prevent the excitatory effects of the isoprostanes (55). In particular, the E-ring isoprostanes 8-iso PGE₁ and 8-iso PGE₂ act at submicromolar concentrations to completely reverse tone elicited by other agonists (e.g., cholinergic stimulation), whereas the F-ring isomers are 10–100-fold less potent.

In a later study, we concluded that the relaxations evoked by 8-iso PGE₂ involve inhibitory EP receptors, because desensitization protocols demonstrated the ability of the isoprostane to attenuate PGE₂ responses and vice versa (17). The subtype of EP receptor involved (EP₂ or EP₄) has not been established due to a lack of commercially available selective antagonists. In the same study, we found that 8-iso PGE₂-evoked relaxations were augmented by rolipram, a selective inhibitor of phosphodiesterase IV, suggesting a key role for cyclic AMP in these responses.

Electrophysiology

Agonist-induced changes in ASM tone are often accompanied by a variety of electrical signaling events, although the role for these is still debated (50). With respect to isoprostanes, there is little published information regarding the ionic events that mediate or accompany isoprostane-induced contraction or relaxation of ASM. We have found that 8-iso PGE, markedly suppresses K+ currents in canine bronchial smooth muscle (17) (Fig. 2) and porcine tracheal smooth muscle, an effect that is normally associated with a contractile response; however, this compound is instead a fairly potent bronchodilator (17, 55). We have recently examined further the receptors underlying the suppression of K⁺ currents by 8-iso PGE, in porcine tracheal smooth muscle; this suppression is mimicked by the TP-receptor agonist U46619 and blocked by the TP antagonist ICI 192605 (unpublished observations), suggesting the involvement of a TP receptor. How-

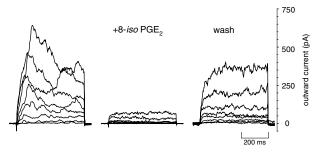


FIG. 2. Isoprostanes suppress potassium currents in dog tracheal smooth muscle. [Catalli *et al.* (17); used by permission.]

ever, U46619 does not itself evoke bronchoconstriction, suggesting these receptors are not present or are not coupled in the usual fashion to the contractile apparatus; instead, these observations might represent further evidence of a novel isoprostane-selective receptor exhibiting cross-reactivity with TP-selective pharmacological agents. K⁺ current suppression is also mimicked by PGE₂. Interestingly, when its suppressive effect on K⁺ currents is prevented by pretreatment with the selective TP-receptor antagonist ICI 192605, 8-iso PGE₂ markedly augments K⁺ currents; this is mimicked only poorly by PGE₂, suggesting a role for some non-TP/non-EP receptor. We have yet to examine the involvement of inhibitory PGD₂-selective prostanoid (DP) and PGI₂-selective prostanoid (IP) receptors in the electrophysiological responses mediated by 8-iso PGE₂.

Augmentation of bronchoconstrictor responses

Isoprostanes may contribute to the nonspecific augmentation of airway responsiveness to various bronchoconstrictors that characterizes a number of airway disorders, including asthma, given that they can augment responsiveness in vascular smooth muscle (99) and in platelets (90). Lipopolysaccharide-induced murine airway hyperresponsiveness is cyclooxygenase (COX)-independent and yet sensitive to a blocker of TP receptors or to the free radical scavenger N-acetylcysteine (37–39), observations that are consistent with the generation and pharmacological actions of isoprostanes. Furthermore, 8iso PGF_{2α} augments methacholine-induced bronchoconstriction in perfused mouse lung in a manner that is sensitive to the TP antagonist SQ 29548 (38). However, this study did not investigate whether isoprostanes and methacholine were acting in an additive or a synergistic fashion; that is, the contractile responses of the isoprostane were not reported, and no mention was made as to whether these responses were inclusive of the agonist/isoprostane combined, and thus may not represent actual hyperresponsiveness.

We recently completed a study in which 8-iso PGE_2 (to a lesser extent, also 8-iso $PGF_{1\alpha}$ and 8-iso $PGF_{2\alpha}$) was found to markedly enhance the sensitivity of bovine tracheal smooth

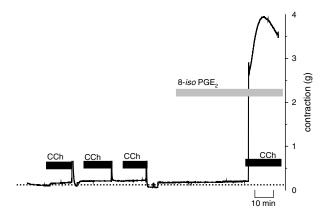


FIG. 3. 8-iso PGE₂ augments responses evoked by a subthreshold concentration of carbachol (CCh; $10^{-9}~M$) in bovine tracheal smooth muscle, without altering tone on its own.

muscle to subthreshold concentrations of carbachol, histamine, and potassium chloride (Fig. 3). This enhancement was far in excess of a mere additive effect, and thus demonstrated a genuine hyperresponsiveness induced by the isoprostane. The hyperresponsiveness was not TP antagonist-sensitive, but was mimicked by the EP and FP receptor agonists PGE_2 and $PGF_{2\alpha}$ (unpublished observations).

Cytokine expression

Interleukin (IL)-1 β stimulates production of several other cytokines (eotaxin, monocyte chemotactic protein) via a pathway in which reactive oxygen species play a key role (126); it may be that isoprostanes mediate this effect of the reactive oxygen species. Consistent with this, in IL-1 β -stimulated cultured human ASM cells, 8-iso PGE₂ (but not 8-iso PGF_{1 α} nor 8-iso PGF_{2 α}) was found to act at submicromolar concentrations to increase the expression of one cytokine [granulocyte colony stimulating factor (G-CSF)], but decrease the expression of another [granulocyte/macrophage colony stimulating factor (GM-CSF)] by 275% and 92%, respectively (20). The inhibitory effect on GM-CSF expression was mediated through EP₂ receptors, whereas the effect on G-CSF involved some other non-TP receptor.

AIRWAY INNERVATION

Neural pathways in the lungs are regulated by prostanoids such as PGE_2 ; this prostanoid sensitizes rat pulmonary vagal C-fibers to mechanical (lung inflation), chemical (capsaicin, lactic acid, adenosine), and electrical stimuli (65), directly activates cholinergic neurons in murine airways (114), and inhibits acetylcholine release from cholinergic nerves in canine airways (136). Given that isoprostanes appear to also stimulate EP receptors, it stands to reason that they might have similar regulatory actions on airway innervation. Indeed, Spicuzza *et al.* (106) demonstrated that 8-*iso* PGF_{2 α} inhibits electrical field stimulation-evoked release of [3H]acetylcholine from parasympathetic nerves of guinea pig airways through some non-TP receptor.

Recently, Clarke et al. (20) examined the effects of two Ering isoprostanes (8-iso PGE, and 8-iso PGE,) in parallel with PGE, and sulprostone (an EP3-selective agonist) on cholinergic neural transmission in guinea-pig trachea. All compounds were found to inhibit electrical field stimulationevoked [3H]acetylcholine release in the rank order PGE, > sulprostone > 8-iso PGE₂ > 8-iso PGE₁. These effects were not reversed by the TP-receptor antagonist SQ 29548 nor the EP₁/EP₂/DP-receptor antagonist AH6809. However, L798106 (a selective EP₃-receptor antagonist) reversed the inhibition of [3H]acetylcholine release by all four compounds. These data suggest that 8-iso PGE1, 8-iso PGE2, PGE2, and sulprostone inhibit cholinergic neurotransmitter release by activating prejunctional EP, receptors. These observations, in conjunction with mechanical data illustrating that isoprostanes directly constrict ASM via TP receptors (17, 55), provide evidence that isoprostanes may play both a protective and deleterious role in airway diseases (106).

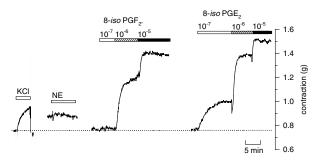


FIG. 4. Isoprostanes evoke contractions in human pulmonary vascular smooth muscle. [Janssen *et al.* (56); used by permission of Nature Publishing Group.]

PULMONARY AND BRONCHIAL VASCULAR SMOOTH MUSCLE

Vasoconstriction

TP receptors are found on virtually every vascular preparation studied, including the pulmonary and bronchial vasculature. It is not surprising, then, that isoprostanes should elicit vasoconstriction in these tissues (Fig. 4) and that those responses should be powerfully blocked by the TP antagonist ICI 192605 (52, 56, 110). 8-iso PGE₂ was by far the most potent and effective of the various isoprostanes we tested (56). Interestingly, in porcine pulmonary vein, this particular isoprostane, but not any of the others, also evoked a contractile response that was resistant to TP-receptor blockade (52). Using a variety of prostanoid agonists and antagonists, we found this response to be exerted through EP receptors (likely of the EP₃ subtype) (52).

We also examined the signaling mechanisms underlying these vasoconstrictor responses (52, 56, 110), finding them to depend on activation of one or more tyrosine kinases, as well as of Rho kinase (consistent with what is known regarding the TP-receptor signaling cascade) (Fig. 5). We did not characterize the specific subtype of tyrosine kinase involved, whether or not it was upstream of Rho kinase, nor the target(s) for these enzymes. Many agonists evoke constriction of smooth muscle via a Rho kinase-mediated phosphorylation of myosin light chain phosphatase, resulting in a net greater phosphorylation of myosin for any given enhancement of the Ca²⁺-dependent myosin light chain kinase activity (a process referred to as increased Ca²⁺ sensitivity of the contractile apparatus).

Using classical fluorimetric techniques in single cells, we found that the isoprostanes generally did not act by releasing internally sequestered Ca²⁺; in fact, the change in [Ca²⁺]_i was generally very small or even absent (unpublished observations). The latter was likely due to voltage-dependent influx of external Ca²⁺, because concurrent patch-clamp electrophysiological studies revealed that isoprostanes suppress K⁺ currents in single pulmonary arterial smooth muscle cells (unpublished observations); this would lead to membrane depolarization and subsequent activation of voltage-dependent Ca²⁺ channels.

Vasodilation

As is the case in ASM, when their effects through TP receptors are blocked, certain isoprostanes can elicit a vasodilatory response in certain vascular beds, including the pulmonary (56) and coronary arteries (134) (Fig. 6); we were unable to find similar vasodilatory effects in cerebral and mesenteric arterial preparations (unpublished observations). The isoprostanes that are able to do this are almost invariably E-ring in nature. It is likely that these effects are mediated through non-TP prostanoid receptor(s), possibly of the EP subtype and/or of the IP subtype. We are currently investigating the latter hypothesis, but this is made difficult by the fact that there are no IP receptor-selective blockers available. Of course, another possibility that needs to be considered is that the isoprostanes are acting through their own unique isoprostane-selective receptors.

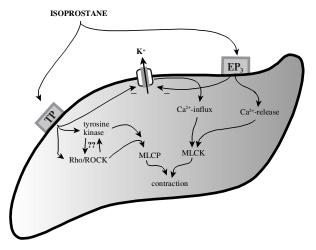


FIG. 5. Summary of the signaling pathways underlying isoprostane-mediated contractions in pulmonary vascular smooth muscle.

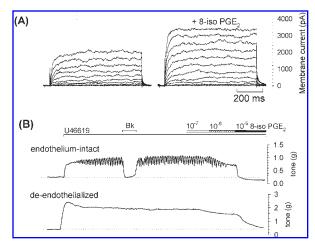


FIG. 6. 8-iso PGE₂ augments potassium currents (A) and elicits endothelium-independent relaxation (B) in porcine coronary vascular smooth muscle. [Zhang et al. (134); used by permission.]

Many vasodilators exert their effects through receptors on the endothelium, which in turn releases relaxant autacoids such as nitric oxide and PGI₂, as well as one or more unidentified molecular species generically referred to as endothelium-derived hyperpolarizing factors (EDHFs). Through the use of inhibitors of nitric oxide synthase and of COX, we excluded any role for nitric oxide or PGI₂ in the isoprostane-evoked responses. Moreover, the relaxations elicited by 8-iso PGE₂ were completely unaffected by removal of the endothelium, suggesting that it acts directly on the smooth muscle rather than indirectly by releasing some EDHF.

While considering the role of the endothelium in mediating the vascular smooth muscle responses to isoprostanes, it occurred to us that isoprostanes themselves might be one of the EDHFs and/or another putative group of endothelium-derived contracting factors (EDCFs), which others have sought for decades to identify (Fig. 7). This hypothesis was based on several lines of evidence.

First, EDHF and EDCF are generally held to be COX-independent metabolites of arachidonic acid (49; 71), a property that is also true of isoprostanes.

Second, EDHF and EDCF have long been associated with reactive oxygen species; their actions are mimicked by peroxide (11, 15, 45, 57, 69, 71, 95, 128, 130, 131), superoxide (40, 60, 120), or hydroxyl radical (89, 96, 97), and can be prevented by free radical scavengers (40, 69, 88, 97). These molecules also play a key role in the generation of isoprostanes.

Third, many describe EDHF responses as being sensitive to inhibitors of phospholipase A_2 (2, 3, 11, 13, 34, 36, 44, 111). This enzyme also plays a key role in releasing the polyunsaturated fatty acid substrates from which the isoprostanes are derived and/or in releasing the isoprostanes after they have been formed within the plasma membrane (78).

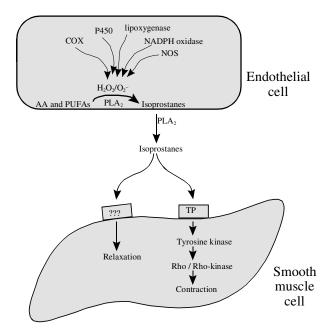


FIG. 7. Hypothesized role of isoprostanes as endotheliumderived hyperpolarizing and contracting factors (EDHF and EDCF, respectively). [Janssen (49); used by permission.]

Fourth, by definition, the endothelium must be able to generate and release EDHF; this too can now be said about isoprostanes (118). Although this has generally been viewed as a result of membrane damage, it is entirely possible that the endothelium might do so in a carefully controlled, enzymatically driven fashion (49). For example, free radicals and reactive oxygen species are produced by COX, cytochrome P450, lipoxygenase, nitric oxide synthase, and NADPH oxidase (25, 29, 69, 112), which in turn are under direct regulation by the endothelial cell (Fig. 7). This could explain some of the reports that EDHF is sensitive to inhibitors of P450 (2, 13, 36), or EDCF to COX inhibitors (129), as well as the apparent insensitivity of EDHF/EDCF to free radical scavengers when they are applied extracellularly (95).

Finally, like EDHF and EDCF, isoprostanes are very powerfully vasoactive, evoking contractions or relaxations in a species-, tissue-, and compound-specific fashion (48, 55, 56).

On the basis of these arguments, we pursued the hypothesis that isoprostanes might represent an EDHF by examining their electrophysiological actions on porcine coronary artery (134) and now more recently in the murine renal artery (unpublished observations). In both tissues, 8-iso PGE₂ elicits substantial augmentation of K⁺ current, membrane hyperpolarization, and vasodilation (Fig. 6). Our data further suggest that the K⁺ channel involved is of the large conductance Ca²⁺-dependent variety (the same type of channel that is activated by EDHF in the porcine coronary artery). The receptors underlying this response are not of the TP subtype, because they persist in the presence of the TP-receptor blocker ICI 192605, but may be of the IP subtype.

AIRWAY EPITHELIUM

Airway epithelial cells assist in the mucociliary trapping and clearance of foreign particles from the airways through the production of mucus, a function that largely relies on salt and water secretion. Of central importance is the secretion of Cl- via the cystic fibrosis transmembrane conductance regulator Cl- channel, whose function is impaired in cystic fibrosis (a disease marked by abnormal mucus production and subsequent malfunction of the mucociliary apparatus). Hydrogen peroxide increases anion transport across monolayers of the human airway epithelial cell line Calu-3 (22); it may be that it does so via production and action of isoprostanes. Consistent with this hypothesis, a recent study found that the isoprostanes 8-iso PGE2 and 8-iso PGF2a do indeed stimulate both apical Cl- and basolateral K+ conductances in the human epithelial cell line Calu-3, with the E-ring isoprostane being more potent and efficacious than the F-ring isoprostane (21). These isoprostanes appear to stimulate anion secretion via an action on TP receptors, because this effect was mimicked by the TP-receptor agonist U46619 and blocked by the TP-receptor antagonist SQ 29548 (21).

INFLAMMATORY CELLS

Inflammation is a complex process, involving adherence of the circulating inflammatory cells to the endothelial cell

wall, their diapedesis across the endothelium *per se*, and migration/chemotaxis toward the site of inflammation, followed by activation of the synthetic and secretory properties of these cells. Recent studies have shown isoprostanes to stimulate or modulate several of these key events.

For example, 8-iso PGE₂ triggers adhesion of monocytes (but not neutrophils) to human endothelial cells via a signaling mechanism that includes stimulation of TP receptors, adenylate cyclase, and protein kinase A (41). Concurrently, 8-iso PGE₂ stimulates phosphorylation of p38 and extracellular signal-regulated kinase (ERK) mitogen-activated protein kinases (41), presumably via protein kinase. Further dissection of the signaling pathway revealed that monocyte adhesion was prevented by inhibition of p38 kinase (but not by inhibition of ERK1/ERK2) and accompanied by increased expression of the early growth response-1 transcription factor, but is independent of the classical nuclear factor-κB (NF-κB) signaling pathway.

In another study, isoprostanes induced expression of IL-8 from human macrophages (102). Here, too, the effect involved activation of p38 and ERK1/2 kinases and increased protein expression (this time, of macrophage inflammatory protein- 1α), but was independent of the NF- κ B pathway.

8-iso PGE_2 and 8-iso $PGF_{2\alpha}$ both enhance human polymorphonuclear granulocyte activity and adhesion to endothelial cells. This effect may be an indirect one, exerted instead via the endothelium, as neither isoprostane increased directly the expression of the membrane marker proteins CD11b or P-selectin, nor were levels of IL-6 or IL-8 altered, but CD11b expression was increased when naive neutrophils were exposed to isoprostane-pretreated endothelial cells or to supernatant of pretreated endothelial cells (132). Interestingly, this indirect activation was not inhibited by antagonists of TP or endothelin receptors. Likewise, the oxidized phospholipid 1-palmitoyl-2-(5,6-epoxyisoprostane E_2)-sn-glycero-3-phosphophocholine induces changes in the expression of cell adhesion molecules of cultured endothelial cells, causing them to adhere monocytes (107, 108, 119).

ISOPROSTANES IN PATHOPHYSIOLOGY

Isoprostanes mediate the effects of free radicals?

It is now widely recognized that inflammation plays a central role in asthma and airway hyperresponsiveness. Inflammatory cells in the airways produce a wide variety of free radicals and reactive oxygen species (including peroxide, superoxide, hypochlorous acid, and hydroxyl radical); these go on to alter many aspects of ASM function, such as contraction of human ASM (91), increase ASM responsiveness in a nonspecific fashion (12, 42), trigger ASM mitogenesis (1), and increase airway epithelial permeability (42, 43, 127). We hypothesized that free radicals and reactive oxygen species exert these effects on ASM in part via peroxidation of membrane lipids and production of isoprostanes. Measured levels of isoprostanes and their metabolites are increased in the plasma, urine, bronchoalveolar lavage fluid, breath condensates, and/or tissues of patients with asthma (63, 73, 125), chronic obstructive pulmonary disease (63, 75), interstitial

lung disease (72), or cystic fibrosis (76), as well as in otherwise normal individuals exposed to ozone (35), cigarette smoke (6, 19, 75, 84), or allergen (74). Consistent with that hypothesis, we found human ASM to contract in response to peroxide (91) and to several isoprostanes (55), whereas canine ASM relaxes in response to peroxide (30, 54) and to Ering isoprostanes (no response to F-ring isoprostanes) (55).

Free radicals also increase membrane permeability in many cell types, and this too may be a consequence of formation of isoprostanes in the membranes by those reactive oxygen species. In particular, the conversion of long-chain polyunsaturated fatty acids (such as arachidonic acid) into a hairpin-like structure (by forming a cyclopentane ring at its center), as well as introducing several hydroxyl groups deep within the membrane (*i.e.*, on the cyclopentane ring), would both be expected to markedly alter membrane fluidity, integrity, and hydrophobicity (77, 78).

Asthma

Many have documented accumulation of 8-iso PGF₂₀ in the lungs in asthma (8, 9, 63, 73, 125). By and large, isoprostanes exert important biological effects on every major cell type found in the lung, and collectively, it may be that these effects culminate in the manifestation of asthma (Fig. 1). For example, it is now known that isoprostanes trigger constriction of the airways (at least in the human lung; see Direct effects on mechanical activity), the bronchial and pulmonary vasculature (see Vasoconstriction), and the lymphatics (85, 86, 105), increase smooth muscle responsiveness (see Augmentation of bronchoconstrictor responses), and augment endothelial permeability. Pulmonary arterial pressures could be further exacerbated by the decreased ventilation and hypoxia associated with bronchoconstriction (via the classic hypoxic pulmonary vasoconstrictor response). The elevated pressure in both the bronchial and pulmonary vasculature combined with the increased endothelial permeability and narrowing of the lymphatics would, in turn, set the stage for edema formation. The isoprostanes may also influence the activities of inflammatory cells (see Inflammatory cells), which are characteristically present in the lungs in asthma, thus exacerbating the bronchoconstriction and edema formation. Over a longer time course, the isoprostanes could contribute to smooth muscle hypertrophy and hyperresponsiveness (both characteristic of asthma and of hypertension).

Pulmonary hypertension

Not only might isoprostanes play a key role in vascular biology as EDHFs and/or EDCFs, there is now good reason to believe that they may be important in vascular pathophysiology. Acute lung injury and pulmonary hypertension are associated with increased metabolism of arachidonic acid (10, 32, 53, 70, 87, 92, 117, 133) and are sensitive to inhibitors of TP receptors (18, 31, 47, 80, 87, 103, 113, 117, 133), thromboxane synthase (18, 53, 80), COX (18, 47, 53, 87, 103), or phospholipase A_2 (66). Whereas one interpretation of these data poses a central role for thromboxane A_2 , recent data linking superoxide and peroxide to these changes (4, 32, 101, 121–124), coupled with the finding that these disease states are accompanied by accumulation of isoprostanes (16, 23,

93), raise another possibility: isoprostanes, generated by COX or nonenzymatically by free radicals, and acting through TP receptors, may mediate these changes. 8-iso PGF₂₀ is released from several sources under conditions associated with acute lung injury and hypertension: from deendothelialized pulmonary artery smooth muscle upon stimulation with growth factors (platelet-derived growth factor, transforming growth factor β), proinflammatory cytokines (tumor necrosis factor- α , interferon- γ , and IL-1 β), peroxide, or superoxide (58, 59, 81); from pulmonary arterial endothelium stimulated with hydrogen peroxide (33); and from renal mesangial cells stimulated with IL-1 (62). Exposure of vascular smooth muscle cells to peroxide causes increased activity of cytosolic phospholipase A₂, accumulation of isoprostanes, expression of preproendothelin mRNA, and production of endothelin-1 (98). Another group showed that stimulation of TP receptors in pulmonary arterial smooth muscle cells by 8-iso PGF₂₀ (or another thromboxane A2 analogue) results in marked production of endothelin-1, and that pulmonary hypertension (as indicated by hypertrophy and increased levels of endothelin-1 and 8-iso PGF₂₀) could be prevented by a TP-receptor blocker (L670596), but not a COX-2 inhibitor (46, 47), suggesting strongly that isoprostanes play a key role in pulmonary hypertension. Isoprostanes may also disrupt endothelial barrier function, trigger pulmonary and systemic vasoconstriction directly (via TP receptors), and/or increase the responsiveness of the smooth muscle to other vasoconstrictors (as described under Augmentation of bronchoconstrictor responses), all of which might contribute further to the hypertension.

Lung transplantation

The finding that isoprostanes powerfully constrict bronchial vasculature is highly clinically relevant, because ischemia–reperfusion injury is associated with generation of isoprostanes (14, 68, 94), a substantial proportion of which may remain esterified within membrane phospholipids and be released over a prolonged period of time. During lung transplantation, then, the donor lungs may represent a major source of isoprostanes for days or weeks after the operation, affecting many parameters of lung function, and possibly ultimately jeopardizing the success of the transplantation *per se* (for which restoration of bronchial blood flow is critical) due to ongoing release of isoprostanes from the oxidatively stressed lungs.

CONCLUSION

In conclusion, isoprostanes have been the subject of investigation for only a little over one decade, and for much of that time they had been viewed primarily as breakdown products of lipid peroxidation. Recently, however, there has been growing interest in their biological actions, particularly in the context of oxidative pathophysiology. As such, they have been elevated from being merely *markers* of oxidative stress to being pathologically relevant *mediators*; perhaps they should even be considered a novel class of inflammatory mediators. Now there is the possibility that isoprostanes may even serve a physiological role in the regulation of vascular

smooth muscle tone as an EDHF. It is clear that there is much to learn about this group of molecules.

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ABBREVIATIONS

ASM, airway smooth muscle; COX, cyclooxygenase; DP, prostaglandin D2-selective prostanoid receptor; EDCF, endothelium-derived contracting factor; EDHF, endothelium-derived hyperpolarizing factor; EP, prostaglandin E2-selective prostanoid receptor; ERK, extracellular signal-regulated kinase; FP, prostaglandin F2-selective prostanoid receptor; GCSF, granulocyte colony stimulating factor; GM-CSF, granulocyte/macrophage colony stimulating factor; IL, interleukin; IP, prostaglandin I2-selective prostanoid receptor; NF-kB, nuclear factor-kB; PG, prostaglandin; TP, thromboxane A2-selective prostanoid receptor.

REFERENCES

- Abe MK, Kartha S, Karpova AY, Li J, Liu PT, Kuo WL, and Hershenson MB. Hydrogen peroxide activates extracellular signal-regulated kinase via protein kinase C, Raf-1, and MEK1. Am J Respir Cell Mol Biol 18: 562–569, 1998.
- Adeagbo AS. Endothelium-derived hyperpolarizing factor: characterization as a cytochrome P450 1A-linked metabolite of arachidonic acid in perfused rat mesenteric prearteriolar bed. *Am J Hypertens* 10: 763–771, 1997.
- Adeagbo AS and Henzel MK. Calcium-dependent phospholipase A2 mediates the production of endothelium-derived hyperpolarizing factor in perfused rat mesenteric prearteriolar bed. *J Vasc Res* 35: 27–35, 1998.
- Amari T, Kubo K, Kobayashi T, and Sekiguchi M. Effects of recombinant human superoxide dismutase on tumor necrosis factor-induced lung injury in awake sheep. J Appl Physiol 74: 2641–2648, 1993.
- Armour CL, Johnson PR, Alfredson ML, and Black JL. Characterization of contractile prostanoid receptors on human airway smooth muscle. *Eur J Pharmacol* 165: 215–222, 1989.
- Bachi A, Zuccato E, Baraldi M, Fanelli R, and Chiabrando C. Measurement of urinary 8-epi-prostaglandin F2alpha, a novel index of lipid peroxidation in vivo, by immunoaffinity extraction/gas chromatography-mass spectrometry. Basal levels in smokers and nonsmokers. Free Radic Biol Med 20: 619–624, 1996.
- 7. Banerjee M, Kang KH, Morrow JD, Roberts LJ, and Newman JH. Effects of a novel prostaglandin, 8-epi-

- PGF2 alpha, in rabbit lung in situ. Am J Physiol 263: H660–H663, 1992.
- 8. Baraldi E, Carraro S, Alinovi R, Pesci A, Ghiro L, Bodini A, Piacentini G, Zacchello F, and Zanconato S. Cysteinyl leukotrienes and 8-isoprostane in exhaled breath condensate of children with asthma exacerbations. *Thorax* 58: 505–509, 2003.
- Baraldi E, Ghiro L, Piovan V, Carraro S, Ciabattoni G, Barnes PJ, and Montuschi P. Increased exhaled 8-isoprostane in childhood asthma. *Chest* 124: 25–31, 2003.
- Barefield ES, Hicks TP, and Philips JB. Thromboxane and pulmonary morphometry in the development of the pulmonary hypertensive response to group B streptococcus. Crit Care Med 22: 506–514, 1994.
- Barlow RS, El Mowafy AM, and White RE. H₂O₂ opens BKCa channels via the PLA₂-arachidonic acid signaling cascade in coronary artery smooth muscle. *Am J Physiol Heart Circ Physiol* 279: H475–H483, 2000.
- Bauer V, Oike M, Tanaka H, Inoue R, and Ito Y. Hydrogen peroxide induced responses of cat tracheal smooth muscle cells. *Br J Pharmacol* 121: 867–874, 1997.
- 13. Bauersachs J, Hecker M, and Busse R. Display of the characteristics of endothelium-derived hyperpolarizing factor by a cytochrome P450-derived arachidonic acid metabolite in the coronary microcirculation. *Br J Pharmacol* 113: 1548–1553, 1994.
- Becker PM, Sanders SP, Price P, and Christman BW. F2-Isoprostane generation in isolated ferret lungs after oxidant injury or ventilated ischemia. *Free Radic Biol Med* 25: 703–711, 1998.
- Beny JL and der Weid PY. Hydrogen peroxide: an endogenous smooth muscle cell hyperpolarizing factor. Biochem Biophys Res Commun 176: 378–384, 1991.
- Carpenter CT, Price PV, and Christman BW. Exhaled breath condensate isoprostanes are elevated in patients with acute lung injury or ARDS. *Chest* 114: 1653–1659, 1998
- Catalli A, Zhang D, and Janssen LJ. Receptors and signaling pathway underlying relaxations to isoprostanes in canine and porcine airway smooth muscle. *Am J Physiol Lung Cell Mol Physiol* 283: L1151–L1159, 2002.
- Cave AC, Manche A, Derias NW, and Hearse DJ. Thromboxane A2 mediates pulmonary hypertension after cardiopulmonary bypass in the rabbit. *J Thorac Cardiovasc* Surg 106: 959–967, 1993.
- Chiabrando C, Valagussa A, Rivalta C, Durand T, Guy A, Zuccato E, Villa P, Rossi JC, and Fanelli R. Identification and measurement of endogenous beta-oxidation metabolites of 8-epi-prostaglandin F2alpha. *J Biol Chem* 274: 1313–1319, 1999.
- Clarke DM, Giembycz MA, Yacoub MH, and Belvisi MG. Regulation of the release of colony stimulating factors from human airway smooth muscle cells by isoprostanes. Am J Respir Crit Care Med 165: A114, 2002.
- Cowley EA. Isoprostane-mediated secretion from human airway epithelial cells. Mol Pharmacol 64: 298–307, 2003.
- 22. Cowley EA and Linsdell P. Oxidant stress stimulates anion secretion from the human airway epithelial cell line Calu-3: implications for cystic fibrosis lung disease. *J Physiol* 543: 201–209, 2002.

- Cracowski JL, Cracowski C, Bessard G, Pepin JL, Bessard J, Schwebel C, Stanke-Labesque F, and Pison C. Increased lipid peroxidation in patients with pulmonary hypertension. *Am J Respir Crit Care Med* 164: 1038– 1042, 2001.
- Elmhurst JL, Betti PA, and Rangachari PK. Intestinal effects of isoprostanes: evidence for the involvement of prostanoid EP and TP receptors. *J Pharmacol Exp Ther* 282: 1198–1205, 1997.
- 25. Fleming I, Michaelis UR, Bredenkotter D, Fisslthaler B, Dehghani F, Brandes RP, and Busse R. Endothelium-derived hyperpolarizing factor synthase (cytochrome P450 2C9) is a functionally significant source of reactive oxygen species in coronary arteries. *Circ Res* 88: 44–51, 2001.
- Fukunaga M, Makita N, Roberts LJ, Morrow JD, Takahashi K, and Badr KF. Evidence for the existence of F2-isoprostane receptors on rat vascular smooth muscle cells. *Am J Physiol* 264: C1619–C1624, 1993.
- Fukunaga M, Takahashi K, and Badr KF. Vascular smooth muscle actions and receptor interactions of 8-isoprostaglandin E2, an E2-isoprostane. *Biochem Biophys Res Commun* 195: 507–515, 1993.
- 28. Fukunaga M, Yura T, Grygorczyk R, and Badr KF. Evidence for the distinct nature of F2-isoprostane receptors from those of thromboxane A2. *Am J Physiol* 272: F477–F483, 1997.
- Fulton D, McGiff JC, Wolin MS, Kaminski P, and Quilley
 J. Evidence against a cytochrome P450-derived reactive
 oxygen species as the mediator of the nitric oxide-independent vasodilator effect of bradykinin in the perfused
 heart of the rat. *J Pharmacol Exp Ther* 280: 702–709,
 1997.
- Gao Y and Vanhoutte PM. Effects of hydrogen peroxide on the responsiveness of isolated canine bronchi: role of prostaglandin E2 and I2. Am J Physiol 263: L402–L408, 1992
- Goff CD, Corbin RS, Theiss SD, Frierson HF, Cephas GA, Tribble CG, Kron IL, and Young JS. Postinjury thromboxane receptor blockade ameliorates acute lung injury. *Ann Thorac Surg* 64: 826–829, 1997.
- Gonzalez PK, Zhuang J, Doctrow SR, Malfroy B, Benson PF, Menconi MJ, and Fink MP. EUK-8, a synthetic superoxide dismutase and catalase mimetic, ameliorates acute lung injury in endotoxemic swine. *J Pharmacol Exp Ther* 275: 798–806, 1995.
- Hart CM, Karman RJ, Blackburn TL, Gupta MP, Garcia JG, and Mohler ER III. Role of 8-epi PGF2alpha, 8-iso-prostane, in H₂O₂-induced derangements of pulmonary artery endothelial cell barrier function. *Prostaglandins Leukot Essent Fatty Acids* 58: 9–16, 1998.
- Hayabuchi Y, Nakaya Y, Matsuoka S, and Kuroda Y. Endothelium-derived hyperpolarizing factor activates Ca²⁺ activated K⁺ channels in porcine coronary artery smooth muscle cells. *J Cardiovasc Pharmacol* 32: 642–649, 1998.
- Hazbun ME, Hamilton R, Holian A, and Eschenbacher WL. Ozone-induced increases in substance P and 8-epiprostaglandin F2 alpha in the airways of human subjects. Am J Respir Cell Mol Biol 9: 568–572, 1993.

36. Hecker M, Bara AT, Bauersachs J, and Busse R. Characterization of endothelium-derived hyperpolarizing factor as a cytochrome P450-derived arachidonic acid metabolite in mammals. *J Physiol* 481 (Pt 2): 407–414, 1994.

- Held HD and Uhlig S. LPS-induced airway hyperreactivity in mice is partly mediated by a non-cyclooxygenase-derived TP-receptor agonist. *Am J Respir Crit Care Med* 159: A868, 1999.
- 38. Held HD and Uhlig S. Mechanisms of endotoxin-induced airway and pulmonary vascular hyperreactivity in mice. *Am J Respir Crit Care Med* 162: 1547–1552, 2000.
- Held HD and Uhlig S. The participation of the cyclooxygenase pathway and oxidative stress in lipopolysaccharide-induced airway- and vascular hyperreactivity in the perfused mouse lung. *Am J Respir Crit Care Med* 161: A591, 2000.
- Hong KW, Rhim BY, Lee WS, Jeong BR, Kim CD, and Shin YW. Release of superoxide-dependent relaxing factor(s) from endothelial cells. *Am J Physiol* 257: H1340– H1346, 1989.
- 41. Huber J, Bochkov VN, Binder BR, and Leitinger N. The isoprostane 8-iso-PGE2 stimulates endothelial cells to bind monocytes via cyclic AMP- and p38 MAP kinase-dependent signaling pathways. *Antioxid Redox Signal* 5: 163–169, 2003.
- Hulsmann AR, Raatgeep HR, den Hollander JC, Stijnen T, Saxena PR, Kerrebijn KF, and de Jongste JC. Oxidative epithelial damage produces hyperresponsiveness of human peripheral airways. Am J Respir Crit Care Med 149: 519–525, 1994.
- 43. Hulsmann AR, Raatgeep HR, den Hollander JC, Bakker WH, Saxena PR, and de Jongste JC. Permeability of human isolated airways increases after hydrogen peroxide and poly-L-arginine. *Am J Respir Crit Care Med* 153: 841–846, 1996.
- 44. Hutcheson IR, Chaytor AT, Evans WH, and Griffith TM. Nitric oxide-independent relaxations to acetylcholine and A23187 involve different routes of heterocellular communication. Role of Gap junctions and phospholipase A2. Circ Res 84: 53–63, 1999.
- Iida Y and Katusic ZS. Mechanisms of cerebral arterial relaxations to hydrogen peroxide. Stroke 31: 2224–2230, 2000.
- 46. Jankov RP, Luo X, Cabacungan J, Belcastro R, Frndova H, Lye SJ, and Tanswell AK. Endothelin-1 and O₂-mediated pulmonary hypertension in neonatal rats: a role for products of lipid peroxidation. *Pediatr Res* 48: 289–298, 2000.
- 47. Jankov RP, Belcastro R, Ovcina E, Lee J, Massaeli H, Lye SJ, and Tanswell AK. Thromboxane A(2) receptors mediate pulmonary hypertension in 60% oxygen-exposed newborn rats by a cyclooxygenase-independent mechanism. Am J Respir Crit Care Med 166: 208–214, 2002.
- Janssen LJ. Isoprostanes: an overview and putative roles in pulmonary pathophysiology. Am J Physiol Lung Cell Mol Physiol 280: L1067–L1082, 2001.
- Janssen LJ. Are endothelium-derived hyperpolarizing and contracting factors isoprostanes? *Trends Pharmacol Sci* 23: 59–62, 2002.

 Janssen LJ. Ionic mechanisms and Ca²⁺ regulation in airway smooth muscle contraction: do the data contradict dogma? *Am J Physiol Lung Cell Mol Physiol* 282: L1161– L1178, 2002.

- Janssen LJ and Daniel EE. Pre- and postjunctional effects of a thromboxane mimetic in canine bronchi. Am J Physiol 261: L271–L276, 1991.
- Janssen LJ and Tazzeo T. Involvement of TP and EP3 receptors in vasoconstrictor responses to isoprostanes in pulmonary vasculature. *J Pharmacol Exp Ther* 301: 1060–1066, 2002.
- Janssens SP, Musto SW, Hutchison WG, Spence C, Witten M, Jung W, and Hales CA. Cyclooxygenase and lipoxygenase inhibition by BW-755C reduces acrolein smoke-induced acute lung injury. *J Appl Physiol* 77: 888–895, 1994.
- 54. Janssen LJ, Netherton SJ, and Walters DK. Ca²⁺-dependent K⁺ channels and Na⁺-K⁺-ATPase mediate H₂O₂- and superoxide-induced relaxations in canine trachealis. *J Appl Physiol* 88: 745–752, 2000.
- Janssen LJ, Premji M, Netherton S, Catalli A, Cox G, Keshavjee S, and Crankshaw DJ. Excitatory and inhibitory actions of isoprostanes in human and canine airway smooth muscle. *J Pharmacol Exp Ther* 295: 506–511, 2000.
- Janssen LJ, Premji M, Netherton S, Coruzzi J, Lu-Chao H, and Cox PG. Vasoconstrictor actions of isoprostanes via tyrosine kinase and Rho kinase in human and canine pulmonary vascular smooth muscles. *Br J Pharmacol* 132: 127–134, 2001.
- Jin N and Rhoades RA. Activation of tyrosine kinases in H₂O₂-induced contraction in pulmonary artery. Am J Physiol 272: H2686–H2692, 1997.
- Jourdan KB, Mitchell JA, and Evans TW. Release of isoprostanes by human pulmonary artery in organ culture: a cyclo-oxygenase and nitric oxide dependent pathway. *Biochem Biophys Res Commun* 233: 668–672, 1997.
- Jourdan KB, Evans TW, Goldstraw P, and Mitchell JA. Isoprostanes and PGE2 production in human isolated pulmonary artery smooth muscle cells: concomitant and differential release. *FASEB J* 13: 1025–1030, 1999.
- Katusic ZS and Vanhoutte PM. Superoxide anion is an endothelium-derived contracting factor. *Am J Physiol* 257: H33–H37, 1989.
- Kawikova I, Barnes PJ, Takahashi T, Tadjkarimi S, Yacoub MH, and Belvisi MG. 8-Epi-PGF2 alpha, a novel noncyclooxygenase-derived prostaglandin, constricts airways in vitro. *Am J Respir Crit Care Med* 153: 590–596, 1996.
- 62. Klein T, Reutter F, Schweer H, Seyberth HW, and Nusing RM. Generation of the isoprostane 8-epi-prostaglandin F2alpha in vitro and in vivo via the cyclooxygenases. *J Pharmacol Exp Ther* 282: 1658–1665, 1997.
- Kostikas K, Papatheodorou G, Ganas K, Psathakis K, Panagou P, and Loukides S. pH in expired breath condensate of patients with inflammatory airway diseases. *Am J Respir Crit Care Med* 165: 1364–1370, 2002.
- 64. Kunapuli P, Lawson JA, Rokach J, and FitzGerald GA. Functional characterization of the ocular prostaglandin

- F2alpha (PGF2alpha) receptor. Activation by the isoprostane, 12-iso-PGF2alpha. *J Biol Chem* 272: 27147–27154, 1997.
- 65. Kwong K and Lee LY. PGE2 sensitizes cultured pulmonary vagal sensory neurons to chemical and electrical stimuli. *J Appl Physiol* 93: 1419–1428, 2002.
- 66. Li HP, He J, Quan CJ, and Ding JS. The relationship between the activity of phospholipase A2 and acute hypoxic pulmonary arterial pressure. *Sheng Li Hsueh Pao* 49: 685–968, 1997.
- 67. MacIntyre DE, Salzman EW, and Gordon JL. Prostaglandin receptors on human platelets. Structure-activity relationships of stimulatory prostaglandins. *Biochem J* 174: 921–929, 1978.
- Mathews WR, Guido DM, Fisher MA, and Jaeschke H. Lipid peroxidation as molecular mechanism of liver cell injury during reperfusion after ischemia. *Free Radic Biol Med* 16: 763–770, 1994.
- 69. Matoba T, Shimokawa H, Nakashima M, Hirakawa Y, Mukai Y, Hirano K, Kanaide H, and Takeshita A. Hydrogen peroxide is an endothelium-derived hyperpolarizing factor in mice. *J Clin Invest* 106: 1521–1530, 2000.
- Mikhail G, Chester AH, Gibbs JS, Borland JA, Banner NR, and Yacoub MH. Role of vasoactive mediators in primary and secondary pulmonary hypertension. *Am J Cardiol* 82: 254–255, 1998.
- Mombouli JV and Vanhoutte PM. Endothelium-derived hyperpolarizing factor(s): updating the unknown. *Trends Pharmacol Sci* 18: 252–256, 1997.
- 72. Montuschi P, Ciabattoni G, Paredi P, Pantelidis P, du Bois RM, Kharitonov SA, and Barnes PJ. 8-Isoprostane as a biomarker of oxidative stress in interstitial lung diseases. *Am J Respir Crit Care Med* 158: 1524–1527, 1998.
- Montuschi P, Corradi M, Ciabattoni G, Nightingale J, Kharitonov SA, and Barnes PJ. Increased 8-isoprostane, a marker of oxidative stress, in exhaled condensate of asthma patients. *Am J Respir Crit Care Med* 160: 216– 220, 1999.
- Montuschi P, Curro D, Ragazzoni E, Preziosi P, and Ciabattoni G. Anaphylaxis increases 8-iso-prostaglandin F2alpha release from guinea-pig lung in vitro. Eur J Pharmacol 365: 59–64, 1999.
- 75. Montuschi P, Collins JV, Ciabattoni G, Lazzeri N, Corradi M, Kharitonov SA, and Barnes PJ. Exhaled 8-isoprostane as an in vivo biomarker of lung oxidative stress in patients with COPD and healthy smokers. *Am J Respir Crit Care Med* 162: 1175–1177, 2000.
- Montuschi P, Kharitonov SA, Ciabattoni G, Corradi M, van Rensen L, Geddes DM, Hodson ME, and Barnes PJ. Exhaled 8-isoprostane as a new non-invasive biomarker of oxidative stress in cystic fibrosis. *Thorax* 55: 205–209, 2000.
- 77. Morrow JD and Roberts LJ. The isoprostanes. Current knowledge and directions for future research. *Biochem Pharmacol* 51: 1–9, 1996.
- Morrow JD, Awad JA, Boss HJ, Blair IA, and Roberts LJ. Non-cyclooxygenase-derived prostanoids (F2-isoprostanes) are formed in situ on phospholipids. *Proc Natl Acad Sci U S A* 89: 10721–10725, 1992.

- 79. Morrow JD, Minton TA, and Roberts LJ. The F2-isoprostane, 8-epi-prostaglandin F2 alpha, a potent agonist of the vascular thromboxane/endoperoxide receptor, is a platelet thromboxane/endoperoxide receptor antagonist. *Prostaglandins* 44: 155–163, 1992.
- Nagata T, Uehara Y, Hara K, Igarashi K, Hazama H, Hisada T, Kimura K, Goto A, and Omata M. Thromboxane inhibition and monocrotaline-induced pulmonary hypertension in rats. *Respirology* 2: 283–289, 1997.
- Natarajan R, Lanting L, Gonzales N, and Nadler J. Formation of an F2-isoprostane in vascular smooth muscle cells by elevated glucose and growth factors. *Am J Physiol* 271: H159–H165, 1996.
- Norel X, Labat C, Gardiner P, and Brink C. Prostanoid contractions in human isolated pulmonary muscle preparations: inhibition by BAY u3405. *Adv Prostaglandin Thromboxane Leukot Res* 21A: 473–476, 1991.
- Norel X, Walch L, Labat C, Gascard JP, Dulmet E, and Brink C. Prostanoid receptors involved in the relaxation of human bronchial preparations. *Br J Pharmacol* 126: 867–872, 1999.
- 84. Obata T, Tomaru K, Nagakura T, Izumi Y, and Kawamoto T. Smoking and oxidant stress: assay of isoprostane in human urine by gas chromatography-mass spectrometry. *J Chromatogr B Biomed Sci Appl* 746: 11–15, 2000.
- Oguogho A, Kaliman J, and Sinzinger H. Levels of eicosanoids (6-oxo-PGF1 alpha and 8-epi-PGF2 alpha) in human and porcine lymphatics and lymph. *Lymphology* 31: 186–189, 1998.
- Oguogho A, Kaliman J, and Sinzinger H. Eicosanoid generation and responsiveness of human lymphatics in hyperlipoproteinemia. *Prostaglandins Leukot Essent Fatty Acids* 62: 47–52, 2000.
- 87. Orr JA, Shams H, Karla W, Peskar BA, and Scheid P. Transient ventilatory responses to endotoxin infusion in the cat are mediated by thromboxane A2. *Respir Physiol* 93: 189–201, 1993.
- 88. Pomposiello S, Rhaleb NE, Alva M, and Carretero OA. Reactive oxygen species: role in the relaxation induced by bradykinin or arachidonic acid via EDHF in isolated porcine coronary arteries. *J Cardiovasc Pharmacol* 34: 567–574, 1999.
- Prasad K and Bharadwaj LA. Hydroxyl radical—a mediator of acetylcholine-induced vascular relaxation. *J Mol Cell Cardiol* 28: 2033–2041, 1996.
- Pratico D, Smyth EM, Violi F, and FitzGerald GA. Local amplification of platelet function by 8-epi prostaglandin F2alpha is not mediated by thromboxane receptor isoforms. *J Biol Chem* 271: 14916–14924, 1996.
- Rabe KF, Dent G, and Magnussen H. Hydrogen peroxide contracts human airways in vitro: role of epithelium. *Am J Physiol* 269: L332–L338, 1995.
- 92. Rautanen M, Gullichsen E, Riutta A, Kuttila K, Mucha I, Nelimarkka O, and Niinikoski J. Experimental fat embolism induces urine 2,3-dinor-6-ketoprostaglandin F1α and 11-dehydrothromboxane B2 excretion in pigs. *Crit Care Med* 25: 1215–1221, 1997.
- Razavi HM, Werhun R, Scott JA, Weicker S, Wang IF, McCormack DG, and Mehta S. Effects of inhaled nitric

oxide in a mouse model of sepsis-induced acute lung injury. *Crit Care Med* 30: 868–873, 2002.

- 94. Reilly MP, Delanty N, Roy L, Rokach J, Callaghan PO, Crean P, Lawson JA, and FitzGerald GA. Increased formation of the isoprostanes IPF2alpha-I and 8-epi-prostaglandin F2alpha in acute coronary angioplasty: evidence for oxidant stress during coronary reperfusion in humans. *Circulation* 96: 3314–3320, 1997.
- Rodriguez-Martinez MA, Garcia-Cohen EC, Baena AB, Gonzalez R, Salaices M, and Marin J. Contractile responses elicited by hydrogen peroxide in aorta from normotensive and hypertensive rats. Endothelial modulation and mechanism involved. *Br J Pharmacol* 125: 1329– 1335, 1998.
- Rosenblum WI. Effects of free radical generation on mouse pial arterioles: probable role of hydroxyl radicals. *Am J Physiol* 245: H139–H142, 1983.
- Rosenblum WI. Hydroxyl radical mediates the endothelium-dependent relaxation produced by bradykinin in mouse cerebral arterioles. Circ Res 61: 601–603, 1987.
- Ruef J, Moser M, Kubler W, and Bode C. Induction of endothelin-1 expression by oxidative stress in vascular smooth muscle cells. *Cardiovasc Pathol* 10: 311–315, 2001.
- Sametz W, Grobuschek T, Hammer-Kogler S, Juan H, and Wintersteiger R. Influence of isoprostanes on vasoconstrictor effects of noradrenaline and angiotensin II. *Eur J Pharmacol* 378: 47–55, 1999.
- 100. Sametz W, Hennerbichler S, Glaser S, Wintersteiger R, and Juan H. Characterization of prostanoid receptors mediating actions of the isoprostanes, 8-iso-PGE(2) and 8-iso-PGF(2alpha), in some isolated smooth muscle preparations. *Br J Pharmacol* 130: 1903–1910, 2000.
- 101. Schnackenberg CG and Wilcox CS. Two-week administration of tempol attenuates both hypertension and renal excretion of 8-iso prostaglandin F2alpha. *Hypertension* 33: 424–428, 1999.
- 102. Scholz H, Yndestad A, Damas JK, Waehre T, Tonstad S, Aukrust P, and Halvorsen B. 8-Isoprostane increases expression of interleukin-8 in human macrophages through activation of mitogen-activated protein kinases. *Cardio*vasc Res 59: 945–954, 2003.
- 103. Schuster DP, Stephenson AH, Holmberg S, and Sandiford P. Effect of eicosanoid inhibition on the development of pulmonary edema after acute lung injury. *J Appl Physiol* 80: 915–923, 1996.
- 104. Sheller JR, Mitchell D, Meyrick B, Oates J, and Breyer R. EP(2) receptor mediates bronchodilation by PGE(2) in mice. *J Appl Physiol* 88: 2214–2218, 2000.
- Sinzinger H, Oguogho A, and Kaliman J. Isoprostane 8epi-prostaglandin F2 alpha is a potent contractor of human peripheral lymphatics. *Lymphology* 30: 155–159, 1997
- 106. Spicuzza L, Barnes PJ, Di Maria GU, and Belvisi MG. Effect of 8-iso-prostaglandin F(2 alpha) on acetylcholine release from parasympathetic nerves in guinea pig airways. Eur J Pharmacol 416: 231–234, 2001.
- 107. Subbanagounder G, Leitinger N, Schwenke DC, Wong JW, Lee H, Rizza C, Watson AD, Faull KF, Fogelman AM, and Berliner JA. Determinants of bioactivity of oxi-

- dized phospholipids. Specific oxidized fatty acyl groups at the sn-2 position. *Arterioscler Thromb Vasc Biol* 20: 2248–2254, 2000.
- 108. Subbanagounder G, Wong JW, Lee H, Faull KF, Miller E, Witztum JL, and Berliner JA. Epoxyisoprostane and epoxycyclopentenone phospholipids regulate monocyte chemotactic protein-1 and interleukin-8 synthesis. Formation of these oxidized phospholipids in response to interleukin-1beta. *J Biol Chem* 277: 7271–7281, 2002.
- 109. Takahashi K, Nammour TM, Fukunaga M, Ebert J, Morrow JD, Roberts LJ, Hoover RL, and Badr KF. Glomerular actions of a free radical-generated novel prostaglandin, 8-epi-prostaglandin F2α, in the rat: evidence for interaction with thromboxane A2 receptors. *J Clin Invest* 90: 136–141, 1992.
- Tazzeo T, Miller J, and Janssen LJ. Vasoconstrictor responses, and underlying mechanisms, to isoprostanes in human and porcine bronchial arterial smooth muscle. *Br J Pharmacol* 140: 759–763, 2003.
- 111. Thangaraju M, Vijayalakshmi T, and Sachdanandam P. Effect of tamoxifen on lipid peroxide and antioxidative system in postmenopausal women with breast cancer. *Cancer* 74: 78–82, 1994.
- Thannickal VJ and Fanburg BL. Reactive oxygen species in cell signaling. Am J Physiol Lung Cell Mol Physiol 279: L1005–L1028, 2000.
- 113. Thies SD, Corbin RS, Goff CD, Binns OA, Buchanan SA, Shockey KS, Frierson HF, Young JS, Tribble CG, and Kron IL. Thromboxane receptor blockade improves oxygenation in an experimental model of acute lung injury. *Ann Thorac Surg* 61: 1453–1457, 1996.
- 114. Tilley SL, Hartney JM, Erikson CJ, Jania C, Nguyen M, Stock J, McNeisch J, Valancius C, Panettieri RA Jr, Penn RB, and Koller BH. Receptors and pathways mediating the effects of prostaglandin E2 on airway tone. Am J Physiol Lung Cell Mol Physiol 284: L599–L606, 2003.
- 115. Ungrin MD, Carriere MC, Denis D, Lamontagne S, Sawyer N, Stocco R, Tremblay N, Metters KM, and Abramovitz M. Key structural features of prostaglandin E(2) and prostanoid analogs involved in binding and activation of the human EP(1) prostanoid receptor. *Mol Pharmacol* 59: 1446–1456, 2001.
- 116. Unmack MA, Rangachari PK, and Skadhauge E. Effects of isoprostanes and prostanoids on porcine small intestine. *J Pharmacol Exp Ther* 296: 434–441, 2001.
- 117. Walmrath D, Pilch J, Scharmann M, Grimminger F, and Seeger W. Severe VA/Q mismatch in perfused lungs evoked by sequential challenge with endotoxin and E. coli hemolysin. J Appl Physiol 76: 1020–1030, 1994.
- 118. Watkins MT, Patton GM, Soler HM, Albadawi H, Humphries DE, Evans JE, and Kadowaki H. Synthesis of 8-epi-prostaglandin F2alpha by human endothelial cells: role of prostaglandin H2 synthase. *Biochem J* 344 (Pt 3): 747–754, 1999.
- 119. Watson AD, Subbanagounder G, Welsbie DS, Faull KF, Navab M, Jung ME, Fogelman AM, and Berliner JA. Structural identification of a novel pro-inflammatory epoxyisoprostane phospholipid in mildly oxidized low density lipoprotein. *J Biol Chem* 274: 24787–24798, 1999.

- 120. Wei EP, Kontos HA, and Beckman JS. Mechanisms of cerebral vasodilation by superoxide, hydrogen peroxide, and peroxynitrite. Am J Physiol 271: H1262–H1266, 1996.
- 121. Weissmann N, Grimminger F, Voswinckel R, Conzen J, and Seeger W. Nitro blue tetrazolium inhibits but does not mimic hypoxic vasoconstriction in isolated rabbit lungs. Am J Physiol 274: L721–L727, 1998.
- 122. Weissmann N, Voswinckel R, Hardebusch T, Rosseau S, Ghofrani HA, Schermuly R, Seeger W, and Grimminger R. Evidence for a role of protein kinase C in hypoxic pulmonary vasoconstriction. *Am J Physiol* 276: L90–L95, 1999.
- 123. Weissmann N, Tadic A, Hanze J, Rose F, Winterhalder S, Nollen M, Schermuly RT, Ghofrani HA, Seeger W, and Grimminger F. Hypoxic vasoconstriction in intact lungs: a role for NADPH oxidase-derived H₂O₂? *Am J Physiol Lung Cell Mol Physiol* 279: L683–L690, 2000.
- 124. Weissmann N, Winterhalder S, Nollen M, Voswinckel R, Quanz K, Ghofrani HA, Schermuly RT, Seeger W, and Grimminger F. NO and reactive oxygen species are involved in biphasic hypoxic vasoconstriction of isolated rabbit lungs. *Am J Physiol Lung Cell Mol Physiol* 280: L638–L645, 2001.
- 125. Wood LG, Fitzgerald DA, Gibson PG, Cooper DM, and Garg ML. Lipid peroxidation as determined by plasma isoprostanes is related to disease severity in mild asthma. *Lipids* 35: 967–974, 2000.
- 126. Wuyts WA, Vanaudenaerde BM, Dupont LJ, Demedts MG, and Verleden GM. N-Acetylcysteine reduces chemokine release via inhibition of p38 MAPK in human airway smooth muscle cells. Eur Respir J 22: 43–49, 2003.
- 127. Yamaya M, Sekizawa K, Masuda T, Morikawa M, Sawai T, and Sasaki H. Oxidants affect permeability and repair of the cultured human tracheal epithelium. *Am J Physiol* 268: L284–L293, 1995.
- 128. Yang Z, Zhang A, Altura BT, and Altura BM. Hydrogen peroxide-induced endothelium-dependent relaxation of rat aorta involvement of Ca²⁺ and other cellular metabolites. *Gen Pharmacol* 33: 325–336, 1999.
- 129. Yang ZH, von Segesser L, Bauer E, Stulz P, Turina M, and Luscher TF. Different activation of the endothelial L-arginine and cyclooxygenase pathway in the human internal

- mammary artery and saphenous vein. Circ Res 68: 52-60, 1991.
- 130. Yang ZW, Zhang A, Altura BT, and Altura BM. Endothelium-dependent relaxation to hydrogen peroxide in canine basilar artery: a potential new cerebral dilator mechanism. *Brain Res Bull* 47: 257–263, 1998.
- 131. Yang ZW, Zheng T, Wang J, Zhang A, Altura BT, and Altura BM. Hydrogen peroxide induces contraction and raises $[Ca^{2+}]_i$ in canine cerebral arterial smooth muscle: participation of cellular signaling pathways. *Naunyn Schmiedebergs Arch Pharmacol* 360: 646–653, 1999.
- 132. Zahler S and Becker BF. Indirect enhancement of neutrophil activity and adhesion to cultured human umbilical vein endothelial cells by isoprostanes (iPF2alpha-III and iPE2-III). *Prostaglandins Other Lipid Mediat* 57: 319–331, 1999.
- 133. Zamora CA, Baron DA, and Heffner JE. Thromboxane contributes to pulmonary hypertension in ischemia– reperfusion lung injury. *J Appl Physiol* 74: 224–229, 1993.
- 134. Zhang Y, Tazzeo T, Hirota S, and Janssen LJ. Vasodilatory and electrophysiological actions of 8-iso-prostaglandin E2 in porcine coronary artery. *J Pharmacol Exp Ther* 305: 1054–1060, 2003.
- Zhang R, Ogletree ML, and Moreland S. Characterization of thromboxane A2/prostaglandin endoperoxide receptors in aorta. *Eur J Pharmacol* 317: 91–96, 1996.
- 136. Zhao WW, Robinson NE, and Yu MF. PGE2 inhibits acetylcholine release from cholinergic nerves in canine but not equine airways. *Prostaglandins Leukot Essent Fatty Acids* 51: 347–355, 1994.

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- 2. Saskia van der Sterren, Eduardo Villamor. 2011. Contractile effects of 15-E2t-isoprostane and 15-F2t-isoprostane on chicken embryo ductus arteriosus. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* **159**:4, 436-444. [CrossRef]
- 3. Judith A. Voynow, Apparao Kummarapurugu. 2011. Isoprostanes and asthma. *Biochimica et Biophysica Acta (BBA) General Subjects*. [CrossRef]
- 4. Ullrich Jahn, Jean-Marie Galano, Thierry Durand. 2008. Jenseits von Prostaglandinen Chemie und Biologie radikalisch gebildeter cyclischer oxygenierter Metabolite von mehrfach ungesättigten Fettsäuren. *Angewandte Chemie* **120**:32, 5978-6041. [CrossRef]
- Ullrich Jahn, Jean-Marie Galano, Thierry Durand. 2008. Beyond Prostaglandins-Chemistry and Biology of Cyclic Oxygenated Metabolites Formed by Free-Radical Pathways from Polyunsaturated Fatty Acids. *Angewandte Chemie International Edition* 47:32, 5894-5955. [CrossRef]
- 6. Yuichiro Sakamoto, Kunihiro Mashiko, Toru Obata, Hisashi Matsumoto, Yoshiaki Hara, Noriyoshi Kutsukata, Yasuhiro Yamamoto. 2008. Relationship Between Treatment Resistance to Hemoperfusion Using a Polymyxin B-Immobilized Fiber Column and Oxidative Stress. ASAIO Journal 54:4, 412-415. [CrossRef]
- 7. Ginger L. Milne, Jason D. MorrowIsoprostanes . [CrossRef]
- 8. SATYAN LAKSHMINRUSIMHA, JAMES A. RUSSELL, ROBIN H. STEINHORN, DANIEL D. SWARTZ, RITA M. RYAN, SYLVIA F. GUGINO, KAREN A. WYNN, VASANTH H. KUMAR, BOBBY MATHEW, KHAVER KIRMANI, FREDERICK C. MORIN. 2007. Pulmonary Hemodynamics in Neonatal Lambs Resuscitated with 21%, 50%, and 100% Oxygen. *Pediatric Research* 62:3, 313-318. [CrossRef]
- 9. Ginger L. Milne, Jason D. Morrow. 2006. Isoprostanes and Related Compounds: Update 2006. *Antioxidants & Redox Signaling* 8:7-8, 1379-1384. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 10. Stéphanie Rolin, Bernard Masereel, Jean-Michel Dogné. 2006. Prostanoids as pharmacological targets in COPD and asthma. *European Journal of Pharmacology* **533**:1-3, 89-100. [CrossRef]
- 11. SATYAN LAKSHMINRUSIMHA, JAMES A. RUSSELL, ROBIN H. STEINHORN, RITA M. RYAN, SYLVIA F. GUGINO, FREDERICK C. MORIN, DANIEL D. SWARTZ, VASANTH H. KUMAR. 2006. Pulmonary Arterial Contractility in Neonatal Lambs Increases with 100% Oxygen Resuscitation. *Pediatric Research* 59:1, 137-141. [CrossRef]
- 12. Jason D. Morrow . 2005. Introduction for Special Forum Issue on Isoprostanes and Related Compounds. *Antioxidants & Redox Signaling* **7**:1-2, 153-156. [Citation] [Full Text PDF] [Full Text PDF with Links]